

# Package ‘BayesianMCPMod’

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**Title** Simulate, Evaluate, and Analyze Dose Finding Trials with Bayesian MCPMod

**Version** 1.3.1

**Description** Bayesian MCPMod (Fleischer et al. (2022) <[doi:10.1002/pst.2193](https://doi.org/10.1002/pst.2193)>) is an innovative method that improves the traditional MCPMod by systematically incorporating historical data, such as previous placebo group data. This package offers functions for simulating, analyzing, and evaluating Bayesian MCPMod trials with normally and binary distributed endpoints. It enables the assessment of trial designs incorporating historical data across various true dose-response relationships and sample sizes. Robust mixture prior distributions, such as those derived with the Meta-Analytic-Predictive approach (Schmidli et al. (2014) <[doi:10.1111/biom.12242](https://doi.org/10.1111/biom.12242)>), can be specified for each dose group. Resulting mixture posterior distributions are used in the Bayesian Multiple Comparison Procedure and modeling steps. The modeling step also includes a weighted model averaging approach (Pinheiro et al. (2014) <[doi:10.1002/sim.6052](https://doi.org/10.1002/sim.6052)>). Estimated dose-response relationships can be bootstrapped and visualized.

**License** Apache License (>= 2)

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<https://github.com/Boehringer-Engelheim/BayesianMCPMod>

**BugReports** <https://github.com/Boehringer-Engelheim/BayesianMCPMod/issues>

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assessDesign	<i>assessDesign</i>
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## Description

This function performs simulation based trial design evaluations for a set of specified dose-response models

### Usage

```

assessDesign(
  n_patients,
  mods,
  prior_list,
  sd = NULL,
  contr = NULL,
  dr_means = NULL,
  data_sim = NULL,
  estimates_sim = NULL,
  n_sim = 1000,
  alpha_crit_val = 0.05,
  modeling = FALSE,
  simple = TRUE,
  avg_fit = TRUE,
  reestimate = FALSE,
  delta = NULL,
  evidence_level = NULL,
  n_bs_samples = 1000,
  med_selection = c("avgFit", "bestFit"),
  probability_scale = FALSE
)

```

### Arguments

n_patients	Vector specifying the planned number of patients per dose group. A minimum of 2 patients are required in each group.
mods	An object of class Mods as specified in the DoseFinding package.
prior_list	A prior_list object specifying the utilized prior for the different dose groups
sd	A positive value, specification of assumed sd. Not required if either data_sim or estimates_sim is provided. Also not required in case of binary endpoint. Default NULL
contr	An object of class optContr as created by the DoseFinding::getContr function. Allows specification of a fixed contrasts matrix. Default NULL.
dr_means	A vector, allows specification of individual (not model based) assumed effects per dose group. Default NULL.
data_sim	An optional data frame for custom simulated data. Must follow the data structure as provided by simulateData(). Default NULL.
estimates_sim	An optional named list of 1) list of vectors for the estimated means per dose group (estimates_sim\$mu_hats) and 2) of list of matrices for the covariance matrices specifying the (estimated) variabilities (estimates_sim\$S_hats). Dimensions of entries must match the number of dose levels. Default NULL.
n_sim	Number of simulations to be performed
alpha_crit_val	(Un-adjusted) Critical value to be used for the MCP testing step. Passed to the getCritProb function for the calculation of adjusted critical values (on the probability scale). Default 0.05.

modeling	Boolean variable defining whether the Mod part of Bayesian MCP-Mod will be performed in the assessment. More heavy on resources. Default FALSE.
simple	Boolean variable defining whether simplified fit will be applied, see <code>?getModelFits</code> . Set automatically to TRUE if argument <code>delta</code> is provided. Passed to <code>getModelFits</code> . Default TRUE.
avg_fit	Boolean variable, defining whether an average fit (based on generalized AIC weights) should be performed in addition to the individual models. Default TRUE.
reestimate	Boolean variable defining whether critical value should be calculated with re-estimated contrasts (see <code>getCritProb</code> function for more details). Default FALSE.
delta	A numeric value for the threshold Delta for the MED assessment. If NULL, no MED assessment is performed. Default NULL.
evidence_level	A numeric value between 0 and 1 for the evidence level gamma for the MED assessment. Only required for Bayesian MED assessment, see <code>?getMED</code> for details. If NULL, MED assessment will be performed on the fitted model according to the argument <code>med_selection</code> . Default NULL.
n_bs_samples	Number of bootstrap samples for the MED assessment if <code>evidence_level</code> is provided. Note that more extreme quantiles (i.e., quantiles closer to 0 or 1) tend to require more bootstrap samples to maintain precision. Default 1000.
med_selection	A string, either "avgFit" or "bestFit", for the method of MED selection. Default "avgFit".
probability_scale	A boolean to specify if the trial has a continuous or a binary outcome. Setting to TRUE will transform calculations from the logit scale to the probability scale, which can be desirable for a binary outcome. Default FALSE.

## Value

Returns success probabilities for the different assumed dose-response shapes, attributes also includes information around average success rate (across all assumed models) and prior Effective sample size.

## Examples

```

mods <- DoseFinding::Mods(linear      = NULL,
                          emax       = c(0.5, 1.2),
                          exponential = 2,
                          betaMod    = c(1, 1),
                          doses      = c(0, 0.5, 2, 4, 8),
                          maxEff     = 6)

sd <- 12
prior_list <- list(Ctrl = RBesT::mixnorm(comp1 = c(w = 1, m = 0, s = 12), sigma = 2),
                  DG_1 = RBesT::mixnorm(comp1 = c(w = 1, m = 1, s = 12), sigma = 2),
                  DG_2 = RBesT::mixnorm(comp1 = c(w = 1, m = 1.2, s = 11), sigma = 2) ,
                  DG_3 = RBesT::mixnorm(comp1 = c(w = 1, m = 1.3, s = 11), sigma = 2) ,
                  DG_4 = RBesT::mixnorm(comp1 = c(w = 1, m = 2, s = 13), sigma = 2))
n_patients <- c(40, 60, 60, 60, 60)

```

```

dose_levels <- c(0, 0.5, 2, 4, 8)

success_probabilities <- assessDesign(
  n_patients = n_patients,
  mods       = mods,
  prior_list = prior_list,
  sd         = sd,
  n_sim      = 1e2) # speed up example run time

success_probabilities

if (interactive()) { # showcasing further functionality

## Analysis with custom data
data_sim <- simulateData(
  n_patients = n_patients,
  dose_levels = dose_levels,
  sd         = sd,
  mods       = mods,
  n_sim      = 10)

success_probabilities_cd <- assessDesign(
  n_patients = n_patients,
  mods       = mods,
  prior_list = prior_list,
  data_sim   = data_sim,
  sd         = sd,
  n_sim      = 1e2) # speed up example run time

success_probabilities_cd

## Analysis with custom dose response relationship
custom_dr_means <- c(1, 2, 3, 4, 5)

success_probs_custom_dr <- assessDesign(
  n_patients = n_patients,
  mods       = mods,
  prior_list = prior_list,
  dr_means   = custom_dr_means,
  sd         = sd,
  n_sim      = 1e2) # speed up example run time

success_probs_custom_dr

## Analysis with custom estimates for means and variabilies
## No simulated data, only simulated model estimates
estimates_sim <- list(mu_hats = replicate(100, list(c(1, 2, 3, 4, 5) + rnorm(5, 0, 1))),
  S_hats = list(diag(1, 5)))

success_probs_custom_est <- assessDesign(
  n_patients = n_patients,
  mods       = mods,
  prior_list = prior_list,

```

```

    estimates_sim = estimates_sim)

success_probs_custom_est

}

if (interactive()) { # takes typically > 5 seconds

# with MED estimation without bootstrapping
# see ?getMED for details

success_probabilities <- assessDesign(
  n_patients    = n_patients,
  mods          = mods,
  prior_list    = prior_list,
  sd            = sd,
  modeling      = TRUE,
  n_sim         = 10, # speed up example run time
  delta         = 7)

  success_probabilities

# with MED estimation with bootstrapping

success_probabilities <- assessDesign(
  n_patients    = n_patients,
  mods          = mods,
  prior_list    = prior_list,
  sd            = sd,
  modeling      = TRUE,
  n_sim         = 10, # speed up example run time
  delta         = 7,
  evidence_level = 0.8)

  success_probabilities

}

```

---

`getBootstrapQuantiles` *getBootstrapQuantiles*

---

### **Description**

A function for the calculation of bootstrapped model predictions. Samples from the posterior distribution are drawn (via the RBesT function `rmix()`) and for every sample the simplified fitting step (see `getModelFits()` function) and a prediction is performed. These fits are then used to identify the specified quantiles. This approach can be considered as the Bayesian equivalent of the frequentist bootstrap approach described in O'Quigley et al. (2017). Instead of drawing  $n$  bootstrap samples from the sampling distribution of the trial dose-response estimates, here the samples are directly taken from the posterior distribution.

**Usage**

```
getBootstrapQuantiles(
  model_fits,
  quantiles,
  n_samples = 1000,
  doses = NULL,
  probability_scale = attr(model_fits, "probability_scale")
)
```

**Arguments**

model_fits	An object of class modelFits, i.e. information about fitted models & corresponding model coefficients as well as the posterior distribution that was the basis for the model fitting
quantiles	A vector of quantiles that should be evaluated
n_samples	Number of samples that should be drawn as basis for the bootstrapped quantiles
doses	A vector of doses for which a prediction should be performed. If NULL, the dose levels of the model_fits will be used. Default NULL.
probability_scale	A boolean variable to specify if the trial has a continuous or a binary outcome. Setting to TRUE will transform predictions from the logit scale to the probability scale, which can be desirable for a binary outcome. Default attr(model_fits, "probability_scale").

**Value**

A tibble with columns for model, dose, and bootstrapped samples

**References**

O'Quigley J, Iasonos A, Bornkamp B. 2017. Handbook of Methods for Designing, Monitoring, and Analyzing Dose-Finding Trials (1st ed.). Chapman and Hall/CRC. doi:10.1201/9781315151984

**Examples**

```
posterior_list <- list(Ctrl = RBest::mixnorm(comp1 = c(w = 1, m = 0, s = 1), sigma = 2),
  DG_1 = RBest::mixnorm(comp1 = c(w = 1, m = 3, s = 1.2), sigma = 2),
  DG_2 = RBest::mixnorm(comp1 = c(w = 1, m = 4, s = 1.5), sigma = 2),
  DG_3 = RBest::mixnorm(comp1 = c(w = 1, m = 6, s = 1.2), sigma = 2),
  DG_4 = RBest::mixnorm(comp1 = c(w = 1, m = 6.5, s = 1.1), sigma = 2))
models <- c("exponential", "linear")
dose_levels <- c(0, 1, 2, 4, 8)
model_fits <- getModelFits(models = models,
  posterior = posterior_list,
  dose_levels = dose_levels,
  simple = TRUE)

bs_quantiles <- getBootstrapQuantiles(model_fits = model_fits,
  quantiles = c(0.025, 0.5, 0.8, 0.975),
```

```

n_samples = 10, # speeding up example run time
doses     = c(0, 6, 8))

bs_quantiles

```

---

```
getBootstrapSamples  getBootstrapSamples
```

---

## Description

A function to return bootstrap samples from the fitted dose-response models. Samples from the posterior distribution are drawn (via the RBeST function `rmix()`) and for every sample the simplified fitting step (see `getModelFits()` function) and a prediction is performed. These samples are returned by this function. This approach can be considered as the Bayesian equivalent of the frequentist bootstrap approach described in O’Quigley et al. (2017). Instead of drawing `n` bootstrap samples from the sampling distribution of the trial dose-response estimates, here the samples are directly taken from the posterior distribution.

## Usage

```

getBootstrapSamples(
  model_fits,
  n_samples = 1000,
  doses = NULL,
  probability_scale = attr(model_fits, "probability_scale")
)

```

## Arguments

<code>model_fits</code>	An object of class <code>modelFits</code> , i.e. information about fitted models & corresponding model coefficients as well as the posterior distribution that was the basis for the model fitting
<code>n_samples</code>	Number of samples that should be drawn
<code>doses</code>	A vector of doses for which a prediction should be performed
<code>probability_scale</code>	A boolean variable to specify if the trial has a continuous or a binary outcome. Setting to <code>TRUE</code> will transform predictions from the logit scale to the probability scale, which can be desirable for a binary outcome. Default <code>attr(model_fits, "probability_scale")</code> .

## Value

A tibble with columns for `sample_id`, `model`, `dose`, `sample`, and `sample_diff`

## References

O’Quigley J, Iasonos A, Bornkamp B. 2017. Handbook of Methods for Designing, Monitoring, and Analyzing Dose-Finding Trials (1st ed.). Chapman and Hall/CRC. doi:10.1201/9781315151984

**Examples**

```

posterior_list <- list(Ctrl = RBesT::mixnorm(comp1 = c(w = 1, m = 0, s = 1), sigma = 2),
  DG_1 = RBesT::mixnorm(comp1 = c(w = 1, m = 3, s = 1.2), sigma = 2),
  DG_2 = RBesT::mixnorm(comp1 = c(w = 1, m = 4, s = 1.5), sigma = 2) ,
  DG_3 = RBesT::mixnorm(comp1 = c(w = 1, m = 6, s = 1.2), sigma = 2) ,
  DG_4 = RBesT::mixnorm(comp1 = c(w = 1, m = 6.5, s = 1.1), sigma = 2))
models          <- c("exponential", "linear")
dose_levels     <- c(0, 1, 2, 4, 8)
model_fits      <- getModelFits(models      = models,
  posterior      = posterior_list,
  dose_levels    = dose_levels,
  simple         = TRUE)

bs_samples <- getBootstrapSamples(model_fits = model_fits,
  n_samples = 10, # speeding up example run time
  doses     = c(0, 6, 8))

bs_samples

```

getContr

*getContr***Description**

This function calculates contrast vectors that are optimal for detecting certain alternatives via applying the function `optContr()` of the `DoseFinding` package. Hereby, 4 different options can be distinguished that are automatically executed based on the input that is provided

1. Bayesian approach: If `dose_weights` and a `prior_list` are provided an optimized contrasts for the posterior sample size is calculated. In detail, in a first step the `dose_weights` (typically the number of patients per dose group) and the prior information is combined by calculating for each dose group a posterior effective sample. Based on this posterior effective sample sizes the allocation ratio is derived, which allows for a calculation on pseudo-optimal contrasts via regular MCPMod are calculated from the regular MCPMod for these specific weights
2. Frequentist approach: If only `dose_weights` are provided optimal contrast vectors are calculated from the regular MCPMod for these specific weights
3. Bayesian approach + re-estimation: If only a `cov_posterior` (i.e. variability of the posterior distribution) is provided, pseudo-optimal contrasts based on these posterior weights will be calculated
4. Frequentist approach+re-estimation: If only a `cov_new_trial` (i.e. the estimated variability of a new trial) is provided, optimal contrast vectors are calculated from the regular MCPMod for this specific covariance matrix.

**Usage**

```

getContr(
  mods,

```

```

dose_levels,
dose_weights = NULL,
prior_list = NULL,
cov_posterior = NULL,
cov_new_trial = NULL
)

```

### Arguments

<code>mods</code>	An object of class 'Mods' as created by the function 'DoseFinding::Mods()'
<code>dose_levels</code>	Vector containing the different dosage levels.
<code>dose_weights</code>	Vector specifying weights for the different doses. Please note that in case this information is provided together with a prior (i.e. Option 1) is planned these two inputs should be provided on the same scale (e.g. patient numbers). Default NULL
<code>prior_list</code>	A list of objects of class 'normMix' as created with 'RBeST::mixnorm()'. Only required as input for Option 1. Default NULL
<code>cov_posterior</code>	A covariance matrix with information about the variability of the posterior distribution, only required for Option 3. Default NULL
<code>cov_new_trial</code>	A covariance matrix with information about the observed variability, only required for Option 4. Default NULL

### Value

An object of class 'optContr' as provided by the function 'DoseFinding::optContr()'.

### Examples

```

dose_levels <- c(0, 0.5, 2, 4, 8)
mods <- DoseFinding::Mods(
  linear      = NULL,
  emax        = c(0.5, 1.2),
  exponential = 2,
  doses       = dose_levels,
  maxEff      = 6)
cov_posterior <- diag(c(2.8, 3, 2.5, 3.5, 4)^2)

contr_mat <- getContr(
  mods      = mods,
  dose_levels = dose_levels,
  cov_posterior = cov_posterior)

```

---

<code>getCritProb</code>	<i>getCritProb</i>
--------------------------	--------------------

---

## Description

This function calculates multiplicity adjusted critical values. The critical values are calculated in such a way that when using non-informative priors the actual error level for falsely declaring a significant trial in the Bayesian MCPMod is controlled (by the specified alpha level). Hereby optimal contrasts of the frequentist MCPMod are applied and two options can be distinguished

1. Frequentist approach: If only `dose_weights` are provided optimal contrast vectors are calculated from the regular MCPMod for these specific weights and the corresponding critical value for this set of contrasts is calculated via the `critVal()` function of the DoseFinding package.
2. Frequentist approach + re-estimation: If only a `cov_new_trial` (i.e. the covariance matrix of a new trial) is provided, optimal contrast vectors are calculated from the regular MCPMod for this specific matrix. Here as well the critical value for this set of contrasts is calculated via the `critVal()` function of the DoseFinding package.

## Usage

```
getCritProb(
  mods,
  dose_levels,
  dose_weights = NULL,
  cov_new_trial = NULL,
  alpha_crit_val = 0.025
)
```

## Arguments

<code>mods</code>	An object of class "Mods" as specified in the DoseFinding package.
<code>dose_levels</code>	Vector containing the different dosage levels.
<code>dose_weights</code>	Vector specifying weights for the different doses, only required for Option i). Default NULL
<code>cov_new_trial</code>	A covariance matrix, only required for Option ii). Default NULL
<code>alpha_crit_val</code>	Significance level. Default set to 0.025.

## Value

Multiplicity adjusted critical value on the probability scale.

## Examples

```
mods <- DoseFinding::Mods(linear      = NULL,
                          emax       = c(0.5, 1.2),
                          exponential = 2,
```

```

                                doses      = c(0, 0.5, 2,4, 8))
dose_levels <- c(0, 0.5, 2, 4, 8)
critVal <- getCritProb(
  mods          = mods,
  dose_weights  = c(50,50,50,50,50), #reflecting the planned sample size
  dose_levels   = dose_levels,
  alpha_crit_val = 0.05)

```

---

getESS

*getESS*


---

### Description

This function calculates the effective sample size for every dose group via `RBesT::ess()`.

### Usage

```
getESS(post_list, method = c("elir", "moment", "morita"), n_digits = 1, ...)
```

### Arguments

<code>post_list</code>	A posterior list object, for which the effective sample size for each dose group should be calculated
<code>method</code>	A string specifying the method of ESS calculation, see <code>?RBesT::ess()</code> .
<code>n_digits</code>	An integer for the number of digits the result should be rounded to.
<code>...</code>	Optional arguments applicable to specific methods, see <code>?RBesT::ess()</code> .

### Value

A vector of the effective sample sizes for each dose group

---

getMED

*getMED*


---

### Description

This function provides information on the minimally efficacious dose (MED). The MED evaluation can either be based on the fitted model shapes (`model_fits`) or on bootstrapped quantiles (`bs_quantiles`).

**Usage**

```
getMED(
  delta,
  evidence_level = 0.5,
  dose_levels = NULL,
  model_fits = NULL,
  bs_quantiles = NULL,
  probability_scale = attr(model_fits, "probability_scale")
)
```

**Arguments**

**delta** A numeric value for the threshold Delta.

**evidence\_level** A numeric value between 0 and 1 for the evidence level gamma. Used for the bs\_quantiles-based evaluation and not used for the model\_fits-based evaluation. Default 0.5.

**dose\_levels** A vector of numerics containing the different dosage levels. Default NULL.

**model\_fits** An object of class modelFits as created with getModelFits(). Default NULL.

**bs\_quantiles** A dataframe created with getBootstrapQuantiles(). Default NULL.

**probability\_scale** A boolean variable to specify if the trial has a continuous or a binary outcome. Setting to TRUE will transform predictions from the logit scale to the probability scale, which can be desirable for a binary outcome. Default attr(model\_fits, "probability\_scale").

**Details**

The function assumes that the 1st dose group is the control dose group.

The bootstrap approach allows for an MED based on decision rules of the form

$$\widehat{\text{MED}} = \arg \min_{d \in \{d_1, \dots, d_k\}} \left\{ \Pr \left( f(d, \hat{\theta}) - f(d_1, \hat{\theta}) > \Delta \right) > \gamma \right\}.$$

The model-shape approach takes the point estimate of the model into account.

**Value**

A matrix with rows for MED reached, MED, and MED index in the vector of dose levels and columns for the dose-response shapes.

**Examples**

```
posterior_list <- list(Ctrl = RBesT::mixnorm(comp1 = c(w = 1, m = 0, s = 1), sigma = 2),
  DG_1 = RBesT::mixnorm(comp1 = c(w = 1, m = 3, s = 1.2), sigma = 2),
  DG_2 = RBesT::mixnorm(comp1 = c(w = 1, m = 4, s = 1.5), sigma = 2),
  DG_3 = RBesT::mixnorm(comp1 = c(w = 1, m = 6, s = 1.2), sigma = 2),
  DG_4 = RBesT::mixnorm(comp1 = c(w = 1, m = 6.5, s = 1.1), sigma = 2))
models <- c("exponential", "linear")
dose_levels <- c(0, 1, 2, 4, 8)
```

```

model_fits    <- getModelFits(models      = models,
                             posterior   = posterior_list,
                             dose_levels = dose_levels,
                             simple     = TRUE)

# MED based on the model_fit:
getMED(delta = 5, model_fits = model_fits)

# MED based on bootstrapped quantiles
bs_quantiles <- getBootstrapQuantiles(model_fits = model_fits,
                                       quantiles  = c(0.025, 0.2, 0.5, 0.8),
                                       n_samples = 100) # speeding up example run time

getMED(delta      = 5,
        evidence_level = 0.8,
        bs_quantiles  = bs_quantiles)

# MED on the probability scale
getMED(delta = 0.1, model_fits = model_fits, probability_scale = TRUE)

```

---

getModelFits

*getModelFits*


---

## Description

Fits dose-response curves for the specified dose-response models, based on the posterior distributions. For the simplified fit, multivariate normal distributions will be approximated and reduced by one-dimensional normal distributions. For the default case, the Nelder-Mead algorithm is used. In detail, for both approaches the mean vector  $\theta^Y$  and the covariance  $\Sigma$  of the (mixture) posterior distributions and the corresponding posterior weights  $\tilde{\omega}_l$  for  $l \in 1, \dots, L$  are used as basis. For the full fit a GLS estimator is used to minimize the following expression for the respective dose-response models  $m$

$$\hat{\theta}_m = \arg \min_{\theta_m} \sum_{l=1}^L \tilde{\omega}_l (\theta_{l_i}^Y - f(\text{dose}_i, \hat{\theta}_m))' \Sigma_l^{-1} (\theta_{l_i}^Y - f(\text{dose}_i, \hat{\theta}_m))$$

Therefore the function `nloptr` of the `nloptr` package is utilized. In the simplified case  $L = 1$ , as the dimension of the posterior is reduced to 1 first. The generalized AIC values are calculated via the formula

$$gAIC_m = \sum_{l=1}^L \tilde{\omega}_l \sum_{i=0}^K \frac{1}{\Sigma_{l_i,i}} (\theta_{l_i}^Y - f(\text{dose}_i, \hat{\theta}_m))^2 + 2p$$

where  $p$  denotes the number of estimated parameters and  $K$  the number of active dose levels. Here as well for the simplified case the formula reduces to one summand as  $L = 1$ . Corresponding `gAIC` based weights for model  $M$  are calculated as outlined in Schorning et al. (2016)

$$\Omega_I(M) = \frac{\exp(-0.5gAIC_M)}{\sum_{m=1}^Q \exp(-0.5gAIC_m)}$$

where  $Q$  denotes the number of models included in the averaging procedure.

**Usage**

```
getModelFits(
  models,
  dose_levels,
  posterior,
  avg_fit = TRUE,
  simple = FALSE,
  probability_scale = FALSE
)
```

**Arguments**

models	A Mods object as created with <code>DoseFinding::Mods()</code> or a vector of model names for which a fit will be performed. Implemented model shapes are "linear", "exponential", "logistic", "emax", "sigEmax", "quadratic", and "betaMod".
dose_levels	A vector containing the different dosage levels.
posterior	A <code>getPosterior</code> object, containing the (multivariate) posterior distribution per dosage level.
avg_fit	Boolean variable, defining whether an average fit (based on generalized AIC weights) should be performed in addition to the individual models. Default TRUE.
simple	Boolean variable, defining whether simplified fit will be applied. Default FALSE.
probability_scale	A boolean variable to specify if the predicted dose-response should be on the logit scale or the probability scale. Setting to TRUE will transform predictions from the logit scale to the probability scale, which can be desirable for a binary outcome. Default FALSE.

**Value**

An object of class `modelFits`. A list containing information about the fitted model coefficients, the prediction per dose group as well as maximum effect and generalized AIC (and corresponding weight) per model.

**References**

Schorning K, Bornkamp B, Bretz F, Dette H. 2016. Model selection versus model averaging in dose finding studies. *Stat Med*; 35; 4021-4040.

**Examples**

```
posterior_list <- list(Ctrl = RBesT::mixnorm(comp1 = c(w = 1, m = 0, s = 1), sigma = 2),
  DG_1 = RBesT::mixnorm(comp1 = c(w = 1, m = 3, s = 1.2), sigma = 2),
  DG_2 = RBesT::mixnorm(comp1 = c(w = 1, m = 4, s = 1.5), sigma = 2),
  DG_3 = RBesT::mixnorm(comp1 = c(w = 1, m = 6, s = 1.2), sigma = 2),
  DG_4 = RBesT::mixnorm(comp1 = c(w = 1, m = 6.5, s = 1.1), sigma = 2))
models <- c("emax", "exponential", "sigEmax", "linear")
dose_levels <- c(0, 1, 2, 4, 8)
```

```

fit      <- getModelFits(models      = models,
                        posterior    = posterior_list,
                        dose_levels  = dose_levels)

fit

fit_simple <- getModelFits(models      = models,
                        posterior    = posterior_list,
                        dose_levels  = dose_levels,
                        simple       = TRUE)

fit_simple

```

---

getPosterior	<i>getPosterior</i>
--------------	---------------------

---

### Description

Either the patient level data or both  $\mu_{\hat{}}$  as well as  $S_{\hat{}}$  must to be provided. If patient level data is provided  $\mu_{\hat{}}$  and  $S_{\hat{}}$  are calculated within the function using a linear model. This function calculates the posterior distribution. Depending on the input for  $S_{\hat{}}$  this step is either performed for every dose group independently via the RBeST function `postmix()` or the `mvpostmix()` function of the `DoseFinding` package is utilized. In the latter case conjugate posterior mixture of multivariate normals are calculated (DeGroot 1970, Bernardo and Smith 1994)

### Usage

```

getPosterior(
  prior_list,
  data = NULL,
  mu_hat = NULL,
  S_hat = NULL,
  calc_ess = FALSE,
  probability_scale = attr(data, "probability_scale")
)

```

### Arguments

<code>prior_list</code>	a prior list with information about the prior to be used for every dose group
<code>data</code>	dataframe containing the information of dose and response. Also a <code>simulateData</code> object can be provided. Default <code>NULL</code> .
<code>mu_hat</code>	vector of estimated mean values (per dose group). Default <code>NULL</code> .
<code>S_hat</code>	covariance matrix specifying the (estimated) variability. The variance-covariance matrix should be provided and the dimension of the matrix needs to match the number of dose groups. Default <code>NULL</code> .

`calc_ess`           boolean variable, indicating whether effective sample size should be calculated. Default FALSE.

`probability_scale`  
A boolean to specify if the trial has a continuous or a binary outcome. Setting to TRUE will transform calculations from the logit scale to the probability scale, which can be desirable for a binary outcome. Default `attr(data, "probability_scale")`.

### Details

Kindly note that one can sample from the `posterior_list` with `lapply(posterior_list, RBest::rmix, n = 10)`.

### Value

`posterior_list`, a posterior list object is returned with information about (mixture) posterior distribution per dose group (more detailed information about the conjugate posterior in case of covariance input for `S_hat` is provided in the attributes)

### References

BERNARDO, JI. M., and Smith, AFM (1994). Bayesian Theory. 81.

### Examples

```
prior_list <- list(
  Ctrl = RBest::mixnorm(comp1 = c(w = 1, m = 0, s = 5), sigma = 2),
  DG_1 = RBest::mixnorm(comp1 = c(w = 1, m = 1, s = 12), sigma = 2),
  DG_2 = RBest::mixnorm(comp1 = c(w = 1, m = 1.2, s = 11), sigma = 2),
  DG_3 = RBest::mixnorm(comp1 = c(w = 1, m = 1.3, s = 11), sigma = 2),
  DG_4 = RBest::mixnorm(comp1 = c(w = 1, m = 2, s = 13), sigma = 2))

mu_hat <- c(0, 1, 1.5, 2, 2.5)
S_hat <- diag(c(5, 4, 6, 7, 8)^2)

posterior_list <- getPosterior(
  prior_list = prior_list,
  mu_hat     = mu_hat,
  S_hat      = S_hat)

summary(posterior_list)
```

**Description**

Performs Bayesian MCP Test step, as described in Fleischer et al. (2022). Tests for a dose-response effect using a model-based multiple contrast test based on the (provided) posterior distribution. In particular for every dose-response candidate the posterior probability is calculated that the contrast is bigger than 0 (based on the posterior distribution of the dose groups). In order to obtain significant test decision we consider the maximum of the posterior probabilities across the different models. This maximum is compared with a (multiplicity adjusted) critical value (on the probability scale).

**Usage**

```
performBayesianMCP(posterior_list, contr, crit_prob_adj)
```

**Arguments**

`posterior_list` An object derived with `getPosterior` with information about the (mixture) posterior distribution per dose group

`contr` An object of class 'optContr' as created by the `getContr()` function. It contains the contrast matrix to be used for the testing step.

`crit_prob_adj` A `getCritProb` object, specifying the critical value to be used for the testing (on the probability scale)

**Value**

Bayesian MCP test result, with information about p-values for the individual dose-response shapes and overall significance

**References**

Fleischer F, Bossert S, Deng Q, Loley C, Gierse J. 2022. Bayesian MCPMod. *Pharmaceutical Statistics*. 21(3): 654-670. doi:10.1002/pst.2193

**Examples**

```
mods <- DoseFinding::Mods(linear      = NULL,
                          emax       = c(0.5, 1.2),
                          exponential = 2,
                          doses      = c(0, 0.5, 2, 4, 8))

dose_levels <- c(0, 0.5, 2, 4, 8)
sd_posterior <- c(2.8, 3, 2.5, 3.5, 4)
contr_mat <- getContr(
  mods          = mods,
  dose_levels   = dose_levels,
  cov_posterior = diag(sd_posterior)^2)
critVal <- getCritProb(
  mods          = mods,
  dose_weights  = c(50, 50, 50, 50, 50), #reflecting the planned sample size
  dose_levels   = dose_levels,
  alpha_crit_val = 0.05)
prior_list <- list(Ctrl = RBesT::mixnorm(comp1 = c(w = 1, m = 0, s = 5), sigma = 2),
                  DG_1 = RBesT::mixnorm(comp1 = c(w = 1, m = 1, s = 12), sigma = 2),
```

```

DG_2 = RBesT::mixnorm(comp1 = c(w = 1, m = 1.2, s = 11), sigma = 2) ,
DG_3 = RBesT::mixnorm(comp1 = c(w = 1, m = 1.3, s = 11), sigma = 2) ,
DG_4 = RBesT::mixnorm(comp1 = c(w = 1, m = 2, s = 13), sigma = 2))
mu <- c(0, 1, 1.5, 2, 2.5)
S_hat <- diag(c(5, 4, 6, 7, 8)^2)
posterior_list <- getPosterior(
  prior_list = prior_list,
  mu_hat      = mu,
  S_hat       = S_hat,
  calc_ess    = TRUE)

performBayesianMCP(posterior_list = posterior_list,
  contr          = contr_mat,
  crit_prob_adj = critVal)

```

---

performBayesianMCPMod *performBayesianMCPMod*

---

### Description

Performs Bayesian MCP Test step and modeling in a combined fashion. See performBayesianMCP() function for MCP Test step and getModelFits() for the modeling step

### Usage

```

performBayesianMCPMod(
  posterior_list,
  contr,
  crit_prob_adj,
  simple = FALSE,
  avg_fit = TRUE,
  delta = NULL,
  evidence_level = NULL,
  med_selection = c("avgFit", "bestFit"),
  n_samples = 1000,
  probability_scale = FALSE
)

```

### Arguments

**posterior\_list** An object of class 'postList' or a list of 'postList' objects as created by getPosterior() containing information about the (mixture) posterior distribution per dose group

**contr** An object of class 'optContr' as created by the getContr() function. It contains the contrast matrix to be used for the testing step.

**crit\_prob\_adj** A getCritProb object, specifying the critical value to be used for the testing (on the probability scale).

simple	Boolean variable, defining whether simplified fit will be applied. Passed to the getModelFits() function. Default FALSE.
avg_fit	Boolean variable, defining whether an average fit (based on generalized AIC weights) should be performed in addition to the individual models. Default TRUE.
delta	A numeric value for the threshold Delta for the MED assessment. If NULL, no MED assessment is performed. Default NULL.
evidence_level	A numeric value between 0 and 1 for the evidence level gamma for the MED assessment. Only required for Bayesian MED assessment, see ?getMED for details. Default NULL.
med_selection	A string, either "avgFit" or "bestFit" based on the lowest gAIC, for the method of MED selection. Default "avgFit".
n_samples	A numerical for the number of bootstrapped samples in case the Bayesian MED assessment is performed. Default 1e3.
probability_scale	A boolean to specify if the trial has a continuous or a binary outcome. Setting to TRUE will transform calculations from the logit scale to the probability scale, which can be desirable for a binary outcome. Default FALSE.

## Value

Bayesian MCP test result as well as modeling result.

## Examples

```

mods <- DoseFinding::Mods(linear      = NULL,
                          emax       = c(0.5, 1.2),
                          exponential = 2,
                          doses      = c(0, 0.5, 2, 4, 8))

dose_levels <- c(0, 0.5, 2, 4, 8)
sd_posterior <- c(2.8, 3, 2.5, 3.5, 4)
contr_mat <- getContr(
  mods          = mods,
  dose_levels   = dose_levels,
  cov_posterior = diag(sd_posterior)^2)
critVal <- getCritProb(
  mods          = mods,
  dose_weights  = c(50, 50, 50, 50, 50), #reflecting the planned sample size
  dose_levels   = dose_levels,
  alpha_crit_val = 0.6) # unreasonable alpha chosen for this example, rather choose 0.05
prior_list <- list(Ctrl1 = RBesT::mixnorm(comp1 = c(w = 1, m = 0, s = 5), sigma = 2),
                  DG_1  = RBesT::mixnorm(comp1 = c(w = 1, m = 1, s = 12), sigma = 2),
                  DG_2  = RBesT::mixnorm(comp1 = c(w = 1, m = 1.2, s = 11), sigma = 2) ,
                  DG_3  = RBesT::mixnorm(comp1 = c(w = 1, m = 1.3, s = 11), sigma = 2) ,
                  DG_4  = RBesT::mixnorm(comp1 = c(w = 1, m = 2, s = 13), sigma = 2))

mu <- c(0, 1, 1.5, 2, 2.5)
S_hat <- diag(c(5, 4, 6, 7, 8)^2)
posterior_list <- getPosterior(
  prior_list = prior_list,

```

```

mu_hat      = mu,
S_hat       = S_hat,
calc_ess    = TRUE)

performBayesianMCPMod(posterior_list = posterior_list,
                      contr          = contr_mat,
                      crit_prob_adj = critVal,
                      simple         = FALSE,
                      delta          = 1.1)

```

---

plot.modelFits

*plot.modelFits*


---

## Description

Plot function based on the `ggplot2` package. Providing visualizations for each model and a average Fit. Black lines show the fitted dose response models and an AIC based average model. Dots indicate the posterior median and vertical lines show corresponding credible intervals (i.e. the variability of the posterior distribution of the respective dose group). To assess the uncertainty of the model fit one can in addition visualize credible bands (default coloring as orange shaded areas). The calculation of these bands is performed via the `getBootstrapQuantiles()` function. The default setting is that these credible bands are not calculated.

## Usage

```

## S3 method for class 'modelFits'
plot(
  x,
  probability_scale = attr(x, "probability_scale"),
  gAIC = TRUE,
  cr_intv = TRUE,
  alpha_CrI = 0.05,
  cr_bands = FALSE,
  alpha_CrB = c(0.05, 0.2, 1),
  n_bs_smpl = 1000,
  acc_color = "orange",
  plot_res = 100,
  plot_diffs = FALSE,
  ...
)

```

## Arguments

`x` An object of type `modelFits`

`probability_scale` A boolean to specify if the trial has a continuous or a binary outcome. Setting to `TRUE` will transform the output from the logit scale to the probability scale, which can be desirable for a binary outcome. Default `attr(x, "probability_scale")`.

gAIC	Logical value indicating whether gAIC values are shown in the plot. Default TRUE
cr_intv	Logical value indicating whether credible intervals are included in the plot. Default TRUE
alpha_CrI	Numerical value of the width of the credible intervals. Default is set to 0.05 (i.e 95% CI are shown).
cr_bands	Logical value indicating whether bootstrapped based credible bands are shown in the plot. Default FALSE
alpha_CrB	Numerical vector of the width of the credible bands. Default is set to $c(0.05, 0.2, 1)$ , i.e, the 95% CB, 80% CB and bootstrapped median are shown.
n_bs_smp1	Number of bootstrap samples being used. Default 1000.
acc_color	Color of the credible bands. Default "orange".
plot_res	Number of plotted doses within the range of the dose levels, i.e., the resolution of the plot. Default 100.
plot_diffs	Logical flag. Plot differences to the control group. Default FALSE.
...	optional parameter to be passed to plot().

### Value

A ggplot2 object

### Examples

```
posterior_list <- list(Ctrl = RBeST::mixnorm(comp1 = c(w = 1, m = 0, s = 1), sigma = 2),
  DG_1 = RBeST::mixnorm(comp1 = c(w = 1, m = 3, s = 1.2), sigma = 2),
  DG_2 = RBeST::mixnorm(comp1 = c(w = 1, m = 4, s = 1.5), sigma = 2) ,
  DG_3 = RBeST::mixnorm(comp1 = c(w = 1, m = 6, s = 1.2), sigma = 2) ,
  DG_4 = RBeST::mixnorm(comp1 = c(w = 1, m = 6.5, s = 1.1), sigma = 2))
models <- c("exponential", "linear", "emax")
dose_levels <- c(0, 1, 2, 4, 8)
model_fits <- getModelFits(models      = models,
  posterior    = posterior_list,
  dose_levels  = dose_levels,
  simple       = TRUE)

plot(model_fits)

# plot with credible bands
plot(model_fits,
  cr_bands      = TRUE,
  plot_diffs    = FALSE,
  probability_scale = FALSE,
  n_bs_smp1     = 1e2) # speeding up example run-time
```

---

predict.modelFits      *predict.modelFits*

---

## Description

This function performs model predictions based on the provided model and dose specifications

## Usage

```
## S3 method for class 'modelFits'
predict(
  object,
  doses = NULL,
  probability_scale = attr(object, "probability_scale"),
  ...
)
```

## Arguments

object	A modelFits object containing information about the fitted model coefficients
doses	A vector specifying the doses for which a prediction should be done
probability_scale	A boolean variable to specify if the trial has a continuous or a binary outcome. Setting to TRUE will transform predictions from the logit scale to the probability scale, which can be desirable for a binary outcome. Default FALSE.
...	Currently without function

## Value

a list with the model predictions for the specified models and doses

## Examples

```
posterior_list <- list(Ctrl = RBesT::mixnorm(comp1 = c(w = 1, m = 0, s = 1), sigma = 2),
  DG_1 = RBesT::mixnorm(comp1 = c(w = 1, m = 3, s = 1.2), sigma = 2),
  DG_2 = RBesT::mixnorm(comp1 = c(w = 1, m = 4, s = 1.5), sigma = 2) ,
  DG_3 = RBesT::mixnorm(comp1 = c(w = 1, m = 6, s = 1.2), sigma = 2) ,
  DG_4 = RBesT::mixnorm(comp1 = c(w = 1, m = 6.5, s = 1.1), sigma = 2))
models <- c("emax", "exponential", "sigEmax", "linear", "betaMod")
dose_levels <- c(0, 1, 2, 4, 8)
fit <- getModelFits(models = models,
  posterior = posterior_list,
  dose_levels = dose_levels)

predict(fit, doses = c(0, 1, 3, 4, 6, 8))
```

---

<code>simulateData</code>	<i>simulateData</i>
---------------------------	---------------------

---

### Description

Function to simulate patient level data for a normally distributed endpoint

### Usage

```
simulateData(
  n_patients,
  dose_levels,
  sd = NULL,
  mods = NULL,
  n_sim = 1000,
  true_model = NULL,
  dr_means = NULL,
  probability_scale = FALSE
)
```

### Arguments

<code>n_patients</code>	Vector containing number of patients as a numerical value per dose-group.
<code>dose_levels</code>	Vector containing the different dosage levels.
<code>sd</code>	Standard deviation on patient level. Can be NULL if <code>probability_scale</code> is TRUE. Default NULL.
<code>mods</code>	An object of class "Mods" as specified in the DoseFinding package. Can be NULL if <code>'dr_means'</code> is not NULL. Default NULL.
<code>n_sim</code>	Number of simulations to be performed, Default is 1000
<code>true_model</code>	A character for model name, e.g. "emax". Assumed true underlying model. If NULL, all dose-response models included in the <code>mods</code> input parameter will be used. Default NULL.
<code>dr_means</code>	an optional vector, with information about assumed effects per dose group. Default NULL.
<code>probability_scale</code>	A boolean to specify if the trial has a continuous or a binary outcome. Setting to TRUE will transform calculations from the logit scale to the probability scale, which can be desirable for a binary outcome. Default FALSE.

### Value

A list object, containing patient level simulated data for all assumed true models. Also providing information about simulation iteration, patient number as well as dosage levels.

**Examples**

```
models <- DoseFinding::Mods(linear      = NULL,
                           linlog     = NULL,
                           emax       = c(0.5, 1.2),
                           exponential = 2,
                           doses      = c(0, 0.5, 2, 4, 8),
                           maxEff     = 6)

dose_levels <- c(0, 0.5, 2, 4, 8)
sd          <- 12
n_patients  <- c(40, 60, 60, 60, 60)

sim_data <- simulateData(n_patients = n_patients,
                        dose_levels = dose_levels,
                        sd          = sd,
                        mods        = models)

head(sim_data)

# custom response "model" shape
custom_dose_response <- c(1, 2, 3, 4, 5)
sim_data_custom_dr   <- simulateData(n_patients = n_patients,
                                    dose_levels = dose_levels,
                                    sd          = sd,
                                    dr_means   = custom_dose_response)

head(sim_data_custom_dr)
```

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